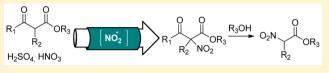
Synthesis of α -Nitro Carbonyls via Nitrations in Flow

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Supporting Information

ABSTRACT: Reported is a safe, rapid method for the synthesis of α -nitro esters via the trapping of nitronium ions. The two-stage nitration and subsequent deacetylation of readily available 1,3-dicarbonyl compounds was achieved using a biphasic semicontinuous approach. α -Nitro esters and amides

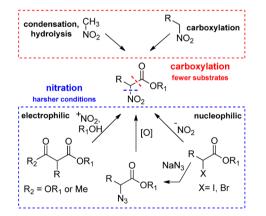


were obtained in good overall yields (53–84%). Some of the α -nitro-1,3-dicarbonyl intermediates exhibit enhanced reactivity and undergo an acid-catalyzed Nef-type reaction to α -oxo-carbonyls.

 α -Nitro esters are valuable synthons in organic synthesis,¹ that can, due to their 1,3-dipole nature and the high acidity of the α proton (p $K_a \sim 5.8$), be further transformed in a variety of ways.² They serve as carbon nucleophiles³ and dipoles for heterocycle synthesis;^{1a,4} they can form phenyliodonium or diazo ylide derivatives,⁵ the latter of which can participate in NH insertion/Mannich-type reactions.⁶ Moreover, α -nitro esters are intermediates for the synthesis of α -keto esters,⁷ γ oxo acids,^{3f} as well α -amino acids.⁵C^{7,8}

Retrosynthetically, the α -nitro ester moiety offers two main disconnections (Scheme 1). The functionalization of a

Scheme 1. Synthetic Approaches for the Synthesis of α -Nitro Esters



nitroalkane with a CO₂ synthon,⁹ is less common. One means of producing the unsubstituted α -nitro ester is the self-condensation of nitromethane under harsh basic conditions followed by an acid-catalyzed esterification.¹⁰ Further alkylations or arylations of this core are possible.^{8c,11}

More commonly, C–N bond formation is the key reaction. The nitro group can be introduced in either a nucleophilic or electrophilic manner. The most direct approach involves the treatment of α -haloesters with a nitrite anion.¹² However, the

substrate scope of this route is limited^{12a,13} and scavengers are required to avoid the formation of α -oximinoesters.^{12b,14} Side product formation can be avoided through a two-step process, transforming α -haloesters via the corresponding α -azidoester (Scheme 1). The desired nitro derivatives are then generated under strong oxidizing conditions (HOF·CH₃CN), rendering this approach less amenable to sensitive substrates, such as olefins and amines.¹⁵

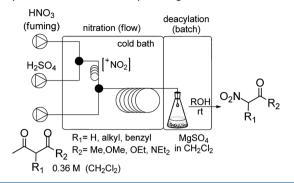
The umpolung approach, where an electrophilic nitronium ion is captured by an enolizable nucleophile, expands the potential pool of starting materials to β -keto esters. Early work by Sifniades provided a homogeneous path, generating the key electrophile intermediate using an acetic anhydride/nitric acid mixture.¹⁶ However, this approach is very temperature sensitive, resulting in side product formation and—disturbingly—the "ejection of the reaction mixture from the reaction vessel".¹⁷

Biphasic systems where a mixture of sulfuric acid and nitric acid (or NH₄NO₃) was added to a chloroform solution of an acetoester at reduced temperatures offered better control over the reaction conditions.^{17,18} The transformation requires vigorous stirring and careful temperature control over the 1 to 3 h it takes to complete. The resulting α -nitroacetate ester derivative is efficiently deacylated to give the nitro ester. While there are few published examples for this process,¹⁶⁻¹⁹ continuous flow nitration of aromatic substrates has allowed for the safer handling of the corrosive strong acids,²⁰ improved temperature control inside the reactor,²¹ and for the prevention of over-nitration.²² In addition, biphasic reactions are significantly accelerated in flow due to the increased interfacial area between the phases. Precise control of the reaction conditions in meso-flow reactors should be a good basis for a general, safe, and broadly applicable process to generate valuable α -nitro esters.

The first step of the nitration/deacylation process (Scheme 2) is electrophile generation. The controlled formation of a

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Scheme 2. Semi-Continuous Setup for the Nitration/ Deacylation of 1,3-Dicarbonyl Compounds



nitronium ion occurred rapidly in a 30 µL PTFE (polytetrafluoroethylene) reactor at 10 °C upon mixing precooled fuming nitric acid (90+%, 1.2 equiv, 0.013 mL min⁻¹) and concentrated sulfuric acid (96%, 6 equiv, 0.080 mL min⁻¹). After a 19 s residence time, the stream of α -acetylbutyrolactone (1) in CH_2Cl_2 (0.36 M, 0.663 mL min⁻¹) was introduced via a T-mixer. The biphasic solution then passed through a second PTFE reactor (0.35 mL, 10 °C, residence time 28 s). The solution was quenched by addition of the exiting stream to a stirred suspension of MgSO₄ in CH₂Cl₂ at room temperature. After completion of the reaction, the quenching agent was filtered off and the solvent removed under reduced pressure. Deacylation occurred by addition and subsequent evaporation of methanol. ¹H NMR analysis revealed the desired α nitrolactone 3 in 63% yield with 92% conversion (Table 1, entry 1).

Both an increase in the residence time using a larger second reactor (0.6 mL, entry 2) as well as the equivalents of nitric acid

Table 1. Optimization of Nitration/Deacylation of α -Acetylbutyrolactone $(1)^a$

_		HNO ₃ , H ₂ SO ₄				$OH O_2 N O O_3$	
entry	HNO ₃ equiv	H ₂ SO ₄ equiv	res. time, (s)	T (°C)	conversion (%) ^b	yield (%) ^b	
1	1.2	6	28 ^c	10	92	63	
2	1.2	6	48	10	> 95	70	
3	1.4	6	47	10	> 95	74	
4	1.4	4	49	10	> 95	70	
5	1.4	7.8	46	10	> 95	79	
6	1.4	10	44	10	> 95	74	
7	1.4 ^d	6.1	47	5	76	60	
8	1.4	7.8	46	15	> 95	74	
9	1.4	7.8	46	5	> 95	75	
10	1.4	7.8	46	0	> 95	73	
11	1.4	7.8	54 ^e	10	> 95	80 (78%) ^f	

^{*a*}Reaction conditions: α -acetylbutyrolactone (1) in CH₂Cl₂ (0.36 M, 0.66 mL min⁻¹); reactor volume 0.6 mL, 96% H₂SO₄ and 90+% fuming HNO₃ were used unless indicated, equivalents with respect to α -acetylbutyrolactone (1); quench: 5 g MgSO₄ in 20 mL CH₂Cl₂, rt; deacylation via addition of 2 mL of methanol. ^{*b*}Determined using mesitylene as internal standard; yield over two steps. ^{*c*}Reactor volume 0.35 mL. ^{*d*}65% Nitric acid. ^{*c*}Reactor volume 0.7 mL. ^{*f*}Isolated yield in parentheses.

(entry 3) resulted in higher yields with complete conversion. While decreasing amounts of sulfuric acid did not improve the yield (70%, entry 4), an increase to 7.8 equiv (entry 5) afforded the desired compound in 79%. Further changes were not advantageous (entry 6). Use of more dilute nitric acid (65%) resulted in the drop in both conversion and yield (entry 7). After additional temperature screenings (entries 8–10), 1.4 equiv nitric acid and 7.8 equiv sulfuric acid at 10 °C were found to be optimal with an overall residence time of 54 s for the nitration (entry 11). The reaction was efficiently quenched with 1 g MgSO₄ per 1 mL acid solution.²³ The productivity of this process, following off-line deacylation, is 1.47 g/hour of the desired 3-nitrodihydrofuran-2(3H)-one (3).

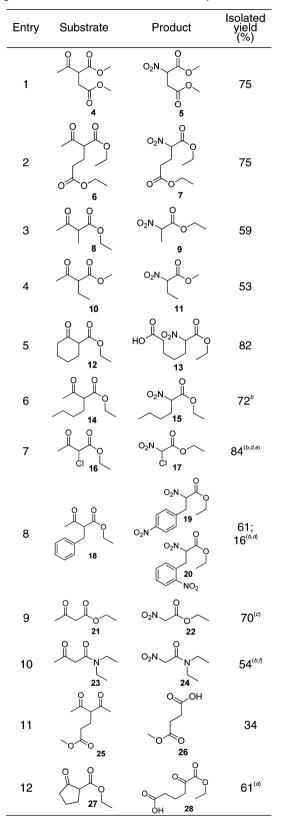
The optimized reaction conditions were tested on a range of 1,3-dicarbonyl compounds (Table 2). α -Substituted acetoacetates bearing an additional ester group gave the corresponding α -nitro esters in good yields (entries 1–2). Alkyl substituted substrates gave moderate [entries 3 (59%), 4 (53%)] to good yields (entry 5, 82%), with the *n*Bu chain requiring slightly higher amounts of acid to achieve full conversion (entry 6, 72%). Electron-withdrawing groups hinder the reaction, with ethyl 2-chloro-3-oxobutanoate (16) necessitating higher temperature (20 °C), a longer reaction time (107 s), and approximately twice as much acid (substrate: 0.2 M, entry 7).²⁴

Several substrates suffer from competing reactions. Ethyl 2benzylacetoacetate (18, entry 8) can also undergo electrophilic aromatic substitution, and the previously optimized conditions resulted in only 71% conversion with multiple substituted products. The reaction was pushed to completion by increasing the equivalents of acid (substrate: 0.16 M). The double nitrated scaffolds **19** (61%) and **20** (16%) were identified as the main products. This represents a limitation of the method, as nitration is believed to occur first at the aromatic ring due to the observation of the solely aryl-nitrated ethyl 2-(4-nitrobenzyl)-3-oxobutanoate (7%).

In the case of α -unsubstituted ethyl acetoacetate **21**, two competing pathways following nitration can occur due to the additional acidic proton: dimerization, which affords a substituted furoxan,¹⁷ and dinitration of the α -position. These pathways could be partly suppressed using a more dilute nitronium ion solution (65% nitric acid) and a larger second reactor (1.8 mL), providing the desired ethyl 2-nitroacetate **22** in 70% yield (entry 9).²⁵

Under the reaction conditions, several α -nitro acyl compounds exhibit enhanced reactivity resulting in the formation of different functional groups. Nitration of *N*,*N*-diethyl-3-oxobutanamide **23** gave a moderate yield of the α -nitro amide (entry 10, 54%) after a longer reaction time (107 s) and a change in the equivalents of nitronium ion (substrate: 0.2 M). One explanation for the lower yield is the formation, post-nitration, of two side products; one resulting from the *in situ* deacylation and dehydration of the nitro group (see Experimental Section for details).

No α -nitro product was observed for the 1,3-diacyl analog methyl 4-acetyl-5-oxohexanoate (25, entry 11). 4-Methoxy-4oxobutanoic acid 26 was isolated as major product (34%). The formation of the carboxylic acid is not surprising as an *in situ* deacylation would result in the formation of a secondary α nitro ketone, which is known to undergo fragmentation under strong acidic conditions to give the corresponding carboxylic acid.²⁶ The α -nitro ketone intermediate can also undergo a Nef reaction, as indicated by the isolation of small amounts both α oxime- and α -oxo-ketone from the same reaction mixture.²⁷ Table 2. Nitration/Deacylation of 1,3-DicarbonylCompounds in a Semi-Continuous Flow System^a



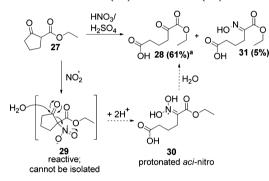
^aStandard nitration conditions: 0.36 M solution in CH₂Cl₂ (0.663 mL min⁻¹); 10 °C; residence time 54 s; HNO₃ (90+%, 0.016 mL min⁻¹), H₂SO₄ (96%, 0.103 mL min⁻¹); for quenching and deacylation procedure see Experimental Section. Yield determined over two steps. ^bHigher amounts of acids used for complete conversion, see

Table 2. continued

Experimental Section. ^c1 M solution in CH_2Cl_2 (0.4 mL min⁻¹), - 5 to -3 °C; residence time 200 s; HNO₃ (65%, 0.031 mL min⁻¹), H_2SO_4 (96%, 0.110 mL min⁻¹). ^dDetermined by NMR using mesitylene as internal standard. ^eResidence time 107 s, 20 °C. ^fResidence time 107 s.

Unexpectedly, this acid-catalyzed Nef reaction—which generally requires a nitronate intermediate—becomes the predominant pathway for ethyl 2-oxocyclopentanecarboxylate 27. Compared to lactone 1, where the α -nitro- γ -lactone is obtained in high yield, the cyclopentanone ester provides α -oxo-ester 28 in 61% yield (entry 12). This result can be explained assuming the formation of the protonated *aci*-nitro species 30 under strong acidic conditions, followed by a Nef reaction to give α -oxo-product 28 and oxime 31—the latter of which was isolated in 5% yield (Scheme 3) and whose formation is known to be dependent on the pH of the reaction medium.²⁸

Scheme 3. Proposed Pathway for the Formation of 6-Ethoxy-5,6-dioxohexanoic Acid (28) and Oxime (31)



"NMR yield using mesitylene as internal standard (47% isolated yield).

In conclusion, a rapid, facile, and safe procedure for the α nitration of 1,3-dicarbonyls via a two-step nitration and deacylation process is disclosed. The controlled nitration in a continuous flow reactor occurs rapidly (54–200 s). Following the quenching of excess H₂SO₄/HNO₃ using MgSO₄, deacylation is achieved in methanol/ethanol in batch. A range of α -nitro esters/amides were produced with moderate-to-good yields (53–84%). Some α -nitro-1,3-dicarbonyl intermediates exhibit enhanced reactivity and under the reaction conditions yielding either Nef α -oxo products or carboxylic acids.

1. EXPERIMENTAL SECTION

General Information. All commercially available compounds and solvents were used without purification. Sulfuric acid (96%) and fuming nitric acid (90+%) were purchased from Roth (ROTIPURAN, 4623.4) and Acros (ACS reagent, A0332793), respectively. Acids and substrates were delivered into the reactor with the help of individual syringe pumps. Both flow reactors were built using polytetrafluoro-ethylene (PTFE) tubing (1.59 mm outer diameter, 0.76 mm inner diameter) and connected by ethylene tetrafluoroethylene (ETFE) T-mixers. All tubing, connectors, and adapters were purchased from IDEX Health and Science. All tubing and mixers were immersed in a water bath, cooled with the help of immersion cooler. Column chromatography was performed using Macherey-Nagel silica gel 60 M (0.04–0.063 mm). Preparative HPLC was performed using a semipreparative YMC-Pack Diol-300-NP column (150 \times 20 mm).

The compounds were visualized by UV (254 nm) and by staining with an aqueous solution of potassium permanganate (prepared from 1.5 g KMnO₄ and 10 g K₂CO₃ in 1.25 mL 10% NaOH in 200 mL water). In describing ¹H and ¹³C NMR spectra the following abbreviations were used to define the multiplicities (s = singlet, d = doublet, dd = doublet of doublets, ddd = doublet of doublet of doublets, t = triplet, dq = doublet of quartets, m = multiplet, br = broad), with coupling constants (J) in Hertz (Hz) and integration. Chemical shifts are reported in parts per million (ppm) relative to residual solvent peaks (δ) and are calibrated to the residual proton and carbon resonance of CDCl₃ (¹H: 7.24, ¹³C: 77.16), CD₃OD (¹H: 3.31, ¹³C: 49.00).²⁹ Highresolution mass spectra were obtained using ESI-Q-TOFmicro mass spectrometer and ESI-TOF mass spectrometer.

General Procedure for Nitration and Deacylation Step. Fuming nitric acid (90+%, 16 µL min⁻¹, 0.343 mmol/min) was mixed with sulfuric acid (96%, 103 µL min⁻¹, 1.85 mmol/min) at 10 °C (cooling bath was used) using a T-mixer. The resulting flow stream was passed through a 0.03 mL PTFE-tubing (0.76 mm inner diameter) and mixed with the solution of the α -acetyl compound (0.36 M in CH₂Cl₂, 0.663 mL min⁻¹, 0.239 mmol/min) using second T-mixer at the same temperature. The biphasic mixture was then passed through a 0.7 mL PTFE-tubing and collected in an Erlenmeyer flask containing a stirred suspension of MgSO₄ (5–10 g) in CH₂Cl₂ (20 mL) at room temperature. MgSO4 was filtered off and the reaction mixture was concentrated under reduced pressure. If deacylation did not occur in situ, the following procedure was performed: the residue was dissolved in 30 mL of the corresponding alcohol (EtOH/MeOH) and stirred from 2 h to overnight (temperature varies from room temperature to reflux depending on the substrate). After completion of the reaction the solvent was removed under reduced pressure and the residue was purified by column chromatography.

3-Nitrodihydrofuran-2(3H)-one (3). General nitration procedure was used (0.6 mL (5.58 mmol) of α -acetylbutyrolactone (1) in CH₂Cl₂ (15 mL)). Quenching and deacylation procedure: for 11 mL of the collected solution, 5 g of MgSO₄ and 20 mL CH₂Cl₂ were used. The reaction mixture was filtered, treated with MeOH (2 mL), and concentrated under reduced pressure (30 °C). The residue was purified by column chromatography (*n*-hexane/ethyl acetate 25:1→ 9:1 ν/ν) to give compound 3 (405 mg, 78%) as an orange oil. R_f = 0.16 (*n*-hexane/ethyl acetate = 3:2 ν/ν); ¹H NMR (400 MHz, CDCl₃): 2.82 (dddd, *J* = 14.0, 9.0, 7.4, 4.8 Hz, 1H), 2.96 (dddd, *J* = 14.4, 8.5, 7.3, 7.3 Hz, 1H), 4.38 (ddd, *J* = 9.2, 7.3, 7.3 Hz, 1H), 4.56 (ddd, *J* = 8.8, 4.9, 4.9 Hz, 1H), 5.38 (dd, *J* = 9.0, 7.4 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃): 27.9, 66.9, 82.2, 167.2; HRMS (ESI): [M + Na]⁺ calcd for C₄H₅NO₄Na: 154.0116, found 154.0106, [M+K]⁺ calcd for C₄H₅NO₄K: 169.9856 found 169.9845.

Dimethyl 2-Nitrosuccinate (5). General nitration procedure was used (0.82 mL (5.05 mmol) of dimethyl 2-acetylsuccinate (4) in CH₂Cl₂ (13.2 mL)). Quenching and deacylation procedure: for 11 mL of the collected solution, 5 g of MgSO₄ and 20 mL CH₂Cl₂ were used, the reaction mixture was filtered and concentrated under reduced pressure. The residue was dissolved in 30 mL MeOH and stirred overnight at room temperature. The solvent was removed under reduced pressure and the residue was purified by column chromatography (*n*-hexane \rightarrow *n*-hexane/ethyl acetate, 100:0 \rightarrow 9:1 *v*/*v*) to give compound 5 (569 mg, 75%) as a yellow oil. R_f = 0.49 (*n*-hexane/ethyl acetate = 5:1 *v*/*v*); ¹H spectrum matches with the literature data.^{30 1}H NMR (400 MHz, CDCl₃): 3.16 (dd, *J* = 17.7, 4.9 Hz, 1H); 3.38 (dd, *J* = 17.7, 9.2 Hz, 1H), 3.75 (s, 3H), 3.57 (dd, *J* = 9.2, 4.9 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃): 3.4.4, 52.8, 54.1, 83.1, 164.2, 168.9. HRMS (ESI): [M+Na]⁺ calcd for C₆H₉NO₆Na: 214.0328, found 214.0322.

Diethyl 2-Nitropentanedioate (7). General nitration procedure was used (1.0 mL (4.65 mmol) of diethyl 2-acetylpentanedioate (6) in CH_2Cl_2 (11.9 mL)). Quenching and deacylation procedure: for 10 mL of the collected solution, 5 g of MgSO₄ and 20 mL CH_2Cl_2 were used, the reaction mixture was filtered and concentrated under reduced pressure. The residue was dissolved in 30 mL EtOH and stirred overnight at room temperature. The solvent was removed under reduced pressure and the residue was purified by column

chromatography (*n*-hexane/ethyl acetate 9:1→2:1 *v*/*v*) to give compound 7 (630 mg, 75%) as a colorless oil. R_f = 0.49 (*n*-hexane/ ethyl acetate = 3:1 *v*/*v*); ¹H spectrum matches with the literature data.³¹ ¹H NMR (400 MHz, CDCl₃): 1.26 (dd, *J* = 7.1, 7.1 Hz, 3H), 1.30 (dd, *J* = 7.1, 7.1 Hz, 3H), 2.39–2.61 (m, 4H), 4.15 (ddd, *J* = 7.1, 7.1, 7.1 Hz, 2H), 4.29 (ddd, *J* = 7.1, 7.1, 7.1 Hz, 2H), 5.27–5.32 (m, 1H); ¹³C NMR (100 MHz, CDCl₃): 14.0, 14.3, 25.4, 29.7, 61.1, 63.3, 86.8, 164.3, 171.6. HRMS (ESI): [M+Na]⁺ calcd for C₉H₁₅NO₆Na: 256.0797, found 256.0809.

Ethyl 2-Nitropropionate (9). General nitration procedure was used (ethyl 2-methylacetoacetate (8) purity 95%, 0.75 mL (5.04 mmol) of substrate 8 in CH₂Cl₂ (13.25 mL)). Quenching and deacylation procedure: for 10 mL of the collected solution, 10 g of MgSO₄ and 20 mL CH₂Cl₂ were used, the reaction mixture was filtered and concentrated under reduced pressure. The residue was dissolved in 30 mL EtOH and stirred for 3 h at 80 °C (temperature of the oil bath). The solvent was removed under reduced pressure and the residue was purified by column chromatography (*n*-hexane/ethyl acetate $15:1\rightarrow10:1 \nu/\nu$) to give compound 9 (311 mg, 59%) as a yellow oil. R_f = 0.43 (*n*-hexane/ethyl acetate = $5:1 \nu/\nu$). Obtained NMR matches with the literature data.³² ¹H NMR (400 MHz, CDCl₃): 1.29 (t, *J* = 7.1 Hz, 3H), 1.77 (d, *J* = 7.2 Hz, 3H), 4.26 (q, *J* = 7.2 Hz, 2H), 5.19 (q, *J* = 7.1 Hz, 1H). ¹³C NMR (100 MHz, CDCl₃): 1.40, 15.8, 63.1, 83.3, 165.2.

Methyl 2-Nitrobutanoate (11). General nitration procedure was used (0.72 mL (5.04 mmol) of methyl 2-ethylacetoacetate (10) in CH₂Cl₂ (13.28 mL)). Quenching and deacylation procedure: for 11 mL of the collected solution, 10 g of MgSO₄ and 20 mL CH₂Cl₂ were used, the reaction mixture was filtered and concentrated under reduced pressure. The residue was dissolved in 30 mL MeOH and stirred for 130 min at 65 °C (temperature of the oil bath). The solvent was removed under reduced pressure and the residue was purified by column chromatography (*n*-hexane/ethyl acetate $10:1 \rightarrow 7:1 \nu/\nu$) to give compound 11 (310 mg, 53%) as a yellow oil. $R_f = 0.45$ (*n*-hexane/ ethyl acetate = 5:1 v/v); ¹H NMR (400 MHz, CDCl₃): 1.04 (dd, J= 7.4, 7.4 Hz, 3H), 2.14–2.36 (m, 2H), 3.83 (s, 3H), 5.05 (dd, J= 9.3, 5.5 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃): 10.3, 24.1, 53.6, 89.4, 165.1; HRMS (ESI): [M+Na]⁺ calcd for C₅H₉NO₄Na: 170.0429, found 170.0419, $[M+K]^+$ calculated for $C_5H_9NO_4K$: 186.0169, found 186.0159. Compound 11 was previously synthesized.9

7-Ethoxy-6-nitro-7-oxoheptanoic Acid (13) and 7-Ethoxy-6-(hydroxyimino)-7-oxoheptanoic Acid (13a). General nitration procedure was used (ethyl 2-oxocyclohexanecarboxylate (12) purity 95%, 0.85 mL (5.05 mmol) of substrate 12 in CH₂Cl₂ (13.15 mL)). Quenching and deacylation procedure: for 4 mL of the collected solution, 5 g of MgSO₄ and 20 mL CH₂Cl₂ were used, the reaction mixture was filtered and concentrated under reduced pressure. The residue was dissolved in 30 mL mixture of EtOH and water (2:1, ν/ν) and stirred overnight at 45 °C (temperature of the oil bath). The solvent was removed under reduced pressure and the residue was purified by column chromatography ($DCM/MeOH 50:1 \rightarrow 5:2 \nu/\nu$) to give compound 13 (275 mg, 82%) as a yellow-green solid and compound 13a (33.4 mg, 11%) as a yellow solid. Analytical data for compound 13: $R_f = 0.30$ (DCM/MeOH = 30:1 ν/ν); mp 57-59 °C; ¹H NMR (400 MHz, CDCl₃):1.29 (t, J = 7.1 Hz, 3H), 1.40–1.49 (m, 2H) 1.66-1.74 (m, 2H), 2.10-2.19 (m, 1H), 2.24-2.32 (m, 1H), 2.36-2.39 (m, 2H), 4.27 (q, J = 7.1 Hz, 2H), 5.09 (dd, J = 9.3, 5.4 Hz, 1H), 9-12 (br signal COOH); ¹³C NMR (100 MHz, CDCl₃) 13.9, 23.8, 25.0, 29.9, 33.5, 63.2, 87.9, 164.5, 179.5; HRMS (ESI): [M+Na]⁺ calcd for C9H15NO6Na 256.0797, found 256.0796. Analytical data for compound 13a: $R_f = 0.5 (DCM/MeOH = 15:1 \nu/\nu); mp 98-100 °C;$ ¹H NMR (400 MHz, $CDCl_3$):1.33 (t, J = 7.1 Hz, 3H), 1.62–1.71 (m, 4H) 2.40 (t, J = 7.1 Hz, 2H), 2.66 (t, J = 7.2 Hz, 2H), 4.28 (q, J = 7.1 Hz, 2H), COOH and N-OH protons are not observed; ¹³C NMR (100 MHz, CDCl₃) 14.1, 24.5, 24.6, 25.3, 33.7, 62.0, 152.0, 163.2, 179.1; HRMS (ESI): [M+Na]⁺ calcd for C₉H₁₅NO₅Na: 240.0848, found 240.0859.

Ethyl 2-Nitrohexanoate (15). Nitric acid (90+%, 16 μ L/min, 0.343 mmol/min) was mixed with sulfuric acid (96%, 103 μ L/min, 1.85 mmol/min) at 10 °C using a T-mixer. The resulting flow stream was

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passed through a 0.03 mL PTFE-tubing and mixed with the ethyl 2acetylhexanoate (14) solution (1.0 mL (5.11 mmol) of substrate 14 in CH₂Cl₂ (15 mL), 0.663 mL/min, 0.212 mmol/min) using second Tmixer at the same temperature. The biphasic mixture was passed through 0.7 mL PTFE-tubing and collected in an Erlenmeyer flask containing stirred suspension of MgSO4 in CH2Cl2 (for 11 mL of the collected solution 5 g of MgSO4 and 20 mL CH2Cl2 were used) at room temperature. The reaction mixture was filtered and concentrated under reduced pressure. The residue was dissolved in 30 mL EtOH and stirred overnight at room temperature. The solvent was removed under reduced pressure and the residue was purified by column chromatography (*n*-hexane \rightarrow *n*-hexane/ethyl acetate, 100:0 \rightarrow 9:1 *v*/*v*) to give compound 15 (480 mg, 72%) as a colorless oil. $R_f = 0.49$ (nhexane/ethyl acetate = 2:1 ν/ν); ¹H NMR (400 MHz, CDCl₃): 0.90 (dd, J = 6.9, 6.9 Hz, 3H); 1.27 (dd, J = 7.1, 7.1 Hz, 3H), 1.32–1.42 (m, 4H), 2.05-2.15 (m, 1H), 2.18-2.31 (m, 1H), 4.25 (ddd, J = 7.1, 7.1, 7.1 Hz, 2H), 5.07 (dd, J = 9.4, 5.5 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃): 13.8, 14.0, 22.1, 27.8, 30.1, 63.1, 88.3, 164.8; HRMS (ESI): [M+Na]⁺ calcd for C₈H₁₅NO₄Na: 212.0899, found 212.0887, [M+K]⁺ calculated for C8H15NO4K: 228.0638, found 228.0630. Compound 15 was previously synthesized.^{15,33}

Ethyl 2-Chloro-2-nitroacetate (17). Nitric acid (90+%, 8 µL/min, 0.171 mmol/min) was mixed with sulfuric acid (96%, 52 μ L/min, 0.923 mmol/min) at 20 $\,^{\circ}\mathrm{C}$ using a T-mixer. The resulting acid mixture was passed through a 0.03 mL PTFE-tubing and mixed with the ethyl 2-chloro-3-oxobutanoate (16) solution (substrate 16 purity 95%, 0.23 mL (1.6 mmol) substrate 16 in CH2Cl2 (7.77 mL), 0.332 mL/min, 0.066 mmol/min) at the same temperature. The biphasic mixture was passed through 0.7 mL PTFE-tubing and collected in an Erlenmeyer flask containing stirred suspension of MgSO₄ in CH₂Cl₂ (for 4 mL of the collected solution 5 g of MgSO₄ and 20 mL CH₂Cl₂ were used) at room temperature. The reaction mixture was filtered and concentrated under reduced pressure. The residue was dissolved in 10 mL EtOH and stirred overnight at room temperature. The solvent was gently removed under reduced pressure (room temperature) to give volatile compound 17 as a yellow oil (84%, determined by ¹H NMR using mesitylene as internal standard). An analytically pure sample was obtained by column chromatography (*n*-hexane/ethyl acetate $20:1 \rightarrow$ 10:1 ν/ν). R_f = 0.60 (*n*-hexane/ethyl acetate = 2:1 ν/ν); Obtained ¹H NMR matches with the literature data.³⁴ ¹H NMR (400 MHz, $CDCl_3$: 1.36 (t, J = 7.2 Hz, 3H), 4.39 (q, J = 6.7 Hz, 2H), 6.23 (s, 1H); ¹³C NMR (100 MHz, CDCl₃) 13.9, 64.9, 85.6, 160.7.

Ethyl 2-Nitro-3-(4-nitrophenyl)propanoate (19), Ethyl 2-Nitro-3-(2-nitrophenyl)propanoate (20), and Ethyl 2-(4-Nitrobenzyl)-3oxobutanoate (20a). Nitric acid (90+%, 16 µL/min, 0.343 mmol/ min) was mixed with sulfuric acid (96%, 103 µL/min, 1.852 mmol/ min) at 10 °C using a T-mixer. The resulting acid mixture was passed through a 0.03 mL PTFE-tubing and mixed with the ethyl 2benzylacetoacetate (18) solution (0.48 mL (2.26 mmol) of substrate 18 in CH₂Cl₂ (13.5 mL), 0.663 mL/min, 0.106 mmol/min) at the same temperature. The biphasic mixture was passed through 0.7 mL PTFE-tubing and collected in an Erlenmeyer flask containing stirred suspension of MgSO₄ in CH₂Cl₂ (for 11 mL of the collected solution, 5 g of MgSO₄ and 20 mL CH₂Cl₂ were used) at room temperature. The reaction mixture was filtered and concentrated under reduced pressure. The residue was dissolved in 30 mL EtOH and stirred overnight at 45 °C. The solvent was removed under reduced pressure to give compound 19 (61%), compound 20 (16%), and compound 20a (7%). (Due to the tedious purification procedure, the yields were determined by ¹H NMR using mesitylene as an internal standard). Analytically pure samples of compound 20 as a yellow oil were obtained using purification by column chromatography (n-hexane/ DCM 2:1 \rightarrow 1:5 ν/ν). Compound **20a** was isolated as a yellow oil from the same column, however was unable to be separated from the impurities according to ¹H NMR. An analytically pure sample of compound 19 was obtained by purification using HPLC Hex/iPrOH (99:1, v/v). Analytical data for compound 19: $R_f = 0.33$ (*n*-hexane/ DCM 1:4 ν/ν); ¹H NMR (400 MHz, CDCl₃):1.30 (t, J = 7.1 Hz, 3H), 3.59 (dd, J = 14.8, 5.3 Hz, 1H), 3.68 (dd, J = 14.8, 9.5 Hz, 1H), 4.31 (q, J = 6.3 Hz, 2H), 5.36 (dd, J = 9.4, 5.7 Hz, 1H), 7.42 (d, J = 8.3 Hz, 100 Hz)

2H), 8.20 (d, J = 8.2 Hz, 2H); ¹³C NMR (100 MHz, CDCl₃): 14.0, 35.9, 63.8, 88.4, 124.4, 130.1, 141.6, 147.8 163.5; HRMS (ESI): [M +Na]⁺ calcd for C₁₁H₁₂N₂O₆Na: 291.0593, found 291.0585. Analytical data for compound **20**: $R_f = 0.63$ (*n*-hexane/DCM 1:4 ν/ν); ¹H NMR (400 MHz, \hat{CDCl}_3): 1.29 (t, J = 6.9 Hz, 3H), 3.71 (dd, J = 14.3, 9.8Hz, 1H), 3.90 (dd, J = 14.4, 4.9 Hz, 1H), 4.31 (q, J = 7.1 Hz, 2H), 5.66 (dd, J = 9.7, 5.0 Hz, 1H), 7.36 (d, J = 7.6 Hz, 1H), 7.50 (dd, J = 7.8, 7.8 Hz, 1H), 7.59 (dd, J = 7.5, 7.5 Hz, 1H), 8.11 (d, J = 8.2 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃): 14.0, 34.4, 63.5, 88.1, 125.9, 129.6, 129.7, 133.5, 134.2, 148.9, 163.8; HRMS-ESI: [M+Na]+ calcd for C11H12N2O6Na 291.0593 found 291.0591. Analytical data for compound 20a: Obtained NMR matches with the literature data.³⁵ $R_f = 0.20 (n-hexane/DCM 1:4 v/v); {}^{1}H NMR (400 MHz, CDCl_3):$ 1.22 (t, J = 7.2 Hz, 3H), 2.24 (s, 3H), 3.25 (t, J = 7.6 Hz, 2H), 3.78 (t, *J* = 8.0, 1H), 4.12–4.21 (m, 2H), 7.36 (d, *J* = 8.8 Hz, 2H), 8.14 (d, *J* = 8.7 Hz, 2H); ¹³C NMR (100 MHz, CDCl₃): 14.2, 29.7, 33.6, 60.8, 62.0, 123.9, 129.9, 146.2, 147.0, 168.6, 201.3.

Ethyl 2-Nitroacetate (22). Nitric acid (65%, 31 µL/min, 0.657 mmol/min) was mixed with sulfuric acid (96%, 110 μ L/min, 1.972 mmol/min) at $-5 \rightarrow -3$ °C using a T-mixer (EtOH/N₂ bath was used, volume of the loop before second T-mixer 0.5 mL). The resulting acid mixture was passed through a 0.03 mL PTFE-tubing and mixed with the ethyl acetoacetate (21) solution (6.5 mL (49 mmol) of substrate 21 in CH₂Cl₂ (42.5 mL), 0.4 mL/min, 0.4 mmol/min) at the same temperature. The biphasic mixture was passed through 1.8 mL PTFEtubing and collected in an Erlenmeyer flask containing stirred suspension of MgSO₄ in CH₂Cl₂ (40.5 mL of solution was collected, 21 g of MgSO4 was used: for 1 mL of the pumped H2SO4 2 g of MgSO₄ was used) at room temperature, the reaction mixture was filtered, dried over Na_2SO_4 , treated with EtOH (1:1 ν/ν) and concentrated under reduced pressure (30 °C). The residue was purified using Kugelrohr distillation at 100 °C, 11 mbar to give compound $\tilde{22}$ (3.8 g, 70%) as a yellow oil. In addition, 22% (determined by NMR using mesitylene as an internal standard) of 3,4bis(ethoxycarbonyl)-1,2,5-oxadiazole 2-oxide (22b)^{4c} was observed. Analytical data for compound 22: ¹H NMR (400 MHz, CDCl₃): 1.28 (t, J = 7.2 Hz, 3H), 4.27 (q, J = 7.1 Hz, 2H), 5.14 (s, 2H); ^{13}C spectrum matches with the literature data. 36 ^{13}C NMR (100 MHz, CDCl₃): 13.9, 63.3, 76.4, 162.1; HRMS (ESI): [M+Na]⁺ calcd for C4H7NO4Na: 156.0273, found 156.0252.

N,N-Diethyl-2-nitroacetamide (24), N,N-Diethyl-2-(hydroxyimino)-3-oxobutanamide (24a), 3,4-Bis(diethylcarbamoyl)-1,2,5-oxadiazole 2-Oxide (24b). Nitric acid (90+%, 8 µL/min, 0.171 mmol/ min) was mixed with sulfuric acid (96%, 52 µL/min, 0.923 mmol/ min) at 10 °C using a T-mixer. The resulting acid mixture was passed through a 0.03 mL PTFE-tubing and mixed with N,N-diethyl-3oxobutanamide (23) solution (0.16 mL (1.01 mmol) of substrate 23 in CH₂Cl₂ (4.84 mL), 0.332 mL/min, 0.066 mmol/min) at the same temperature. The biphasic mixture was passed through 0.7 mL PTFEtubing and collected in an Erlenmeyer flask containing stirred suspension of MgSO₄ in CH₂Cl₂ (for 4 mL of the collected solution, 5 g of MgSO₄ and 20 mL CH_2Cl_2 were used) at room temperature. The reaction mixture was filtered and concentrated under reduced pressure. Crude reaction mixture was purified by column chromatography (n-hexane/ethyl acetate $5:1 \rightarrow 1:1 \ v/v$) to give compound 24 (68.7 mg, 54%) as a yellow oil, compound 24b (23.4 mg, 10%) as a yellow oil and compound 24a (16.7 mg, 11%) as a light peach clear solid. Analytical data for compound 24: $R_f = 0.19$ (*n*-hexane/ethyl acetate = 2:1 ν/ν); ¹H NMR (400 MHz, CDCl₃): 1.17 (t, J = 7.1 Hz, 3H), 1.23 (t, J = 7.1 Hz, 3H), 3.24 (q, J = 7.2 Hz, 2H), 3.44 (q, J = 7.1 Hz, 2H), 5.27 (s, 2H); ¹³C NMR (100 MHz, CD₃OD): 12.9, 14.2, 42.0, 43.3, 78.3, 163.2; HRMS (ESI): [M+Na]⁺ calcd for C₆H₁₂N₂O₃Na: 183.0746, found 183.0740. Analytical data for compound 24a: $R_f = 0.07$ (*n*-hexane/ethyl acetate = 2:1 ν/ν); Obtained NMR matches with literature data.³⁷ ¹H NMR (400 MHz, CDCl₃): 1.12 (t, J = 7.0 Hz, 3H), 1.22 (t, J = 7.6 Hz, 3H), 2.39 (s, 3H), 3.12 (q, J = 6.7 Hz, 2H), 3.52 (q, J = 7.0 Hz, 2H), 11.64 (s, 1H); ¹³C NMR (100 MHz, CDCl₃): 12.7, 14.0, 25.7, 39.3, 42.9, 152.4, 164.2, 195.2. Analytical data for compound 24b: $R_f = 0.31$ (*n*-hexane/ ethyl acetate =2:1 ν/ν); ¹H NMR (400 MHz, CDCl₃): 1.18–1.32 (m,

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12H), 3.31 (q, J = 7.2 Hz, 2H), 3.48–3.55 (m, 4H), 3.59 (q, J = 7.1 Hz, 2H); ¹³C NMR (100 MHz, CDCl₃): 12.6, 12.7, 14.4, 14.7, 40.5, 41.1, 43.3., 43.8, 111.5, 152.3, 154.8, 156.8; HRMS (ESI): [M+Na]⁺ calcd for C₁₂H₂₀N₄O₄Na: 307.1382, found 307.1372, [M+K]⁺ calcd for C₁₂H₂₀N₄O₄K: 323.1122, found 323.1111.

4-Methoxy-4-oxobutanoic Acid (26), Methyl 4,5-Dioxohexanoate (26a), Methyl 4-(Hydroxyimino)-5-oxohexanoate (26b). General nitration procedure was used (0.38 mL (2.16 mmol) of substrate 25 in CH₂Cl₂ (5.62 mL)). Quenching and deacylation procedure: for 4 mL of the collected solution, 5 g of MgSO4 and 20 mL CH2Cl2 were used, the reaction mixture was filtered and concentrated under reduced pressure. Crude was purified by column chromatography (n-hexane/ ethyl acetate 5:1 \rightarrow 1:1 and then DCM \rightarrow DCM/MeOH 100:1 \rightarrow $60:1 \rightarrow 5:1 \nu/\nu$ to give compound **26** (65.4 mg, 34%) as an yellow oil together with compound 26a (5.6 mg, 3%) as a yellow oil, and compound 26b (22.0 mg, 9%) as a white amorphous solid. Additionally unreacted starting material was identified by ¹H NMR but was not isolated due to tedious purification procedure. Analytical data for compound 26: Obtained NMR matches with the literature data.³⁸ ¹H NMR (400 MHz, CDCl₃): 2.60-2.69 (m, 4H), 3.69 (s, 3H), 8.5–9.5 (br, COOH); ¹³C NMR (100 MHz, CDCl₃): 28.8, 29.1, 52.1, 172.8, 178.4. Analytical data for compound 26a: R_f = 0.4 (nhexane/ethyl acetate =4:1 ν/ν); ¹H NMR (600 MHz, CDCl₃): 2.36 (s, 3H), 2.66 (t, J = 6 Hz 2H), 3.03 (t, J = 6 Hz, 2H), 3.69 (s, 3H); ¹³C NMR (151 MHz, CDCl₃): 23.8, 27.8, 30.9, 52.1, 172.9, 197.1, 197.6; HRMS (ESI): [M+Na]⁺ calcd for C₇H₁₀O₄Na: 181.0477, found 181.0478. Compound 26a was previously synthesized.³⁹ Analytical data for compound **26b**: $R_f = 0.37$ (DCM/MeOH = 30:1 ν/ν); ¹H NMR (400 MHz, CDCl₃): 2.37 (s, 3H), 2.52 (t, J = 7.8 Hz, 2H), 2.84 (t, J = 7.7 Hz, 2H), 3.67 (s, 3H), 7.89 (s, 1H); ¹³C NMR (100 MHz, CDCl₃): 18.2, 25.4, 30.2, 52.0, 158.8, 173.2, 196.7; HRMS (ESI): [M +Na]⁺ calcd for C₇H₁₁NO₄Na: 196.0586, found 196.0579, [M+K]⁺ calcd for C7H11NO4K 212.0325, found 212.0322.

6-Ethoxy-5,6-dioxohexanoic Acid (28), 6-Ethoxy-5-(hydroxyimino)-6-oxohexanoic Acid (31), 6-Ethoxy-5-nitro-6-oxohexanoic Acid (28b). General nitration procedure was used (0.75 mL (5.06 mmol) of ethyl 2-oxocyclopentanecarboxylate (27) in CH₂Cl₂ (13.25 mL)). Quenching and deacylation procedure: for 11 mL of the collected solution, 10 g of MgSO₄ and 20 mL CH₂Cl₂ were used, the reaction mixture was filtered and concentrated under reduced pressure. Analysis of crude ¹H NMR revealed formation of 6-ethoxy-5,6dioxohexanoic acid (28) with the 61% yield determined by ¹H NMR using mesitylene as internal standard. Presumably ethyl 1-nitro-2oxocyclopentanecarboxylate (29) was formed, as suggested by the ¹H NMR spectrum (¹H NMR (400 MHz, CDCl₃): 2.05-2.13 (m, 2H), 2.53–2.61 (m, 2H), 2.81 (dt, J = 14.5, 7.3 Hz, 1H), 2.96 (dt, J = 14.0, 6.8 Hz, 1H), peaks corresponding to EtO-group are overlapping with EtO-group of another compound) which, upon standing, spontaneously undergoes a ring-opening reaction with atmospheric water to give compound 28b. The residue was purified by column chromatography (*n*-hexane/ethyl acetate $15:1 \rightarrow 10:1$ and then DCM/MeOH 60:1 \rightarrow 15:1 ν/ν) to give compound 28 (353 mg, 47%) as a yellow oil, oxime 31 (42.2 mg, 5%) as a white solid and compound 28b (33 mg, 4%) as a white solid. Analytical data for compound 28: $R_f = 0.45$ (DCM/MeOH = 15:1 $\nu/\nu/\nu$); Analytical data for the compound 28 matches with the literature data.40 1H NMR (400 MHz, CDCl₃): 1.37 (t, J = 7.1 Hz, 3H), 1.97 (quin, J = 7.2 Hz, 2H), 2.44 (t, J = 7.2 Hz, 2H), 2.95 (t, J = 7.1 Hz, 2H), 4.32 (q, J = 7.1 Hz, 2H), 9–11 (br signal COOH); ¹³C NMR (100 MHz, CDCl₃): 14.0, 17.9, 32.6, 38.2, 62.6, 160.8, 179.2, 193.7. Analytical data for compound 31: $R_f = 0.31$ (DCM/MeOH = 15:1 $\nu/\nu/\nu$); mp 106–108 °C; ¹H NMR: (400 MHz, CDCl₃): 1.35 (t, J = 7.1 Hz, 3H), 1.96 (p, J = 6.9 Hz, 2H), 2.45 (t, J = 6.9 Hz, 2H), 2.73 (t, J = 7.0 Hz, 2H), 4.30 (q, J = 7.1 Hz, 2H), 10-12 (br signal COOH), signal for N-OH proton is not observed; ¹³C NMR (100 MHz, CDCl₃) 14.1, 21.0, 24.3, 33.6, 62.1, 151.9, 163.2, 178.2; HRMS (ESI): [M+Na]⁺ calcd for C₈H₁₃NO₅Na 226.0691, found 226.0686. Analytical data for compound **28b**: $R_f = 0.48$ (DCM/MeOH = 15:1 $\nu/\nu/\nu$); mp 47–49 °C; ¹H NMR (400 MHz, CDCl₃): 1.31 (dd, J = 7.1, 7.1 Hz, 3H), 1.67-1.80 (m, 2H), 2.18-2.27 (m, 1H), 2.29-2.39 (m, 1H), 2.46 (t, J

= 7.1 Hz, 2H), 4.29 (ddd, *J* = 7.1, 7.1, 7.1 Hz, 2H), 5.12 (dd, *J* = 9.3, 5.5 Hz, 1H), 10–12 (br signal COOH); 13 C NMR (100 MHz, CDCl₃) 14.0, 20.8, 29.5, 33.0, 63.3, 87.8, 164.4, 178.7; HRMS (ESI): [M-H]⁺ calcd for C₈H₁₂NO₆: 218.0665, found 218.0686.

ASSOCIATED CONTENT

S Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acs.joc.6b01634.

¹H and ¹³C NMR spectra for all new compounds and optimization experiments to determine minimum amount of desiccant needed for the quenching procedure (PDF)

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Notes

The authors declare no competing financial interest.

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REFERENCES

(1) (a) Kislyi, V. V.; Samet, A. V.; Semenov, V. V. Curr. Org. Chem. 2001, 5, 553–570. (b) Tietze, L. F.; Schneider, C. In Encyclopedia of Reagents for Organic Synthesis; John Wiley & Sons, Ltd, 2001. (c) Shipchandler, M. Synthesis 1979, 1979, 666–686.

(2) Goumont, R.; Magnier, E.; Kizilian, E.; Terrier, F. J. Org. Chem. 2003, 68, 6566-6570.

(3) (a) Ji, C.-B.; Liu, Y.-L.; Cao, Z.-Y.; Zhang, Y.-Y.; Zhou, J. *Tetrahedron Lett.* 2011, *52*, 6118–6121. (b) Luzzio, F. A. *Tetrahedron* 2001, *57*, 915–945. (c) Parise, L.; Pelagalli, A.; Pellacani, L.; Sciubba, F.; Vergari, M. C.; Fioravanti, S. J. Org. Chem. 2016, *81*, 2864–2874. (d) Zhang, S.; Xu, K.; Guo, F.; Hu, Y.; Zha, Z.; Wang, Z. Chem. - Eur. J. 2014, *20*, 979–982. (e) Zhou, Y.; Liu, Q.; Gong, Y. *Tetrahedron Lett.* 2013, *54*, 3011–3014. (f) Aginagalde, M.; Bello, T.; Masdeu, C.; Vara, Y.; Arrieta, A.; Cossio, F. P. J. Org. Chem. 2010, *75*, 7435–7438. (g) Lin, S.; Li, M.; Dong, Z.; Liang, F.; Zhang, J. Org. Biomol. Chem. 2014, *12*, 1341–1350. (h) Dauzonne, D.; Royer, R. Synthesis 1983, 1983, 836–837. (i) Xu, J.; Ai, J.; Liu, S.; Peng, X.; Yu, L.; Geng, M.; Nan, F. Org. Biomol. Chem. 2014, *12*, 3721–3734.

(4) (a) Gangadhara Chary, R.; Rajeshwar Reddy, G.; Ganesh, Y. S. S.; Vara Prasad, K.; Raghunadh, A.; Krishna, T.; Mukherjee, S.; Pal, M. Adv. Synth. Catal. 2014, 356, 160–164. (b) Chen, K.-P.; Chen, Y.-J.; Chuang, C.-P. Eur. J. Org. Chem. 2010, 2010, 5292–5300. (c) Trogu, E.; Cecchi, L.; De Sarlo, F.; Guideri, L.; Ponticelli, F.; Machetti, F. Eur. J. Org. Chem. 2009, 2009, 5971–5978. (d) Guideri, L.; De Sarlo, F.; Machetti, F. Chem. - Eur. J. 2013, 19, 665–677. (e) Baranov, M. S.; Yampolsky, I. V. Tetrahedron Lett. 2013, 54, 628–629. (f) Nakamura, S.; Sugimoto, H.; Ohwada, T. J. Am. Chem. Soc. 2007, 129, 1724– 1732.

(5) (a) Xu, X.; Lu, H.; Ruppel, J. V.; Cui, X.; Lopez de Mesa, S.; Wojtas, L.; Zhang, X. P. J. Am. Chem. Soc. 2011, 133, 15292-15295.
(b) Moreau, B.; Alberico, D.; Lindsay, V. N. G.; Charette, A. B. Tetrahedron 2012, 68, 3487-3496. (c) Moreau, B.; Charette, A. B. J. Am. Chem. Soc. 2005, 127, 18014-18015. (d) Charette, A. B.; Wurz, R. P.; Ollevier, T. Helv. Chim. Acta 2002, 85, 4468-4484. (e) Charette, A. B.; Wurz, R. J. Mol. Catal. A: Chem. 2003, 196, 83-91.

The Journal of Organic Chemistry

(7) Metz, A. E.; Kozlowski, M. C. J. Org. Chem. 2013, 78, 717–722.
(8) (a) Young, A. J.; White, M. C. J. Am. Chem. Soc. 2008, 130, 14090–14091. (b) Chen, Z.; Morimoto, H.; Matsunaga, S.; Shibasaki, M. J. Am. Chem. Soc. 2008, 130, 2170–2171. (c) Singh, A.; Yoder, R. A.; Shen, B.; Johnston, J. N. J. Am. Chem. Soc. 2007, 129, 3466–3467. (d) He, L.; Srikanth, G. S. C.; Castle, S. L. J. Org. Chem. 2005, 70, 8140–8147. (e) Singh, A.; Johnston, J. N. J. Am. Chem. Soc. 2008, 130, 5866–5867. (f) Ram, S.; Ehrenkaufer, R. E. Synthesis 1986, 1986, 133–135.

(9) (a) Reddy, G. R.; Mukherjee, D.; Chittoory, A. K.; Rajaram, S. Org. Lett. **2014**, 16, 5874–5877. (b) Lehr, F.; Gonnermann, J.; Seebach, D. Helv. Chim. Acta **1979**, 62, 2258–2275.

(10) (a) Zen, S.; Koyama, M.; Koto, S. In Org. Synth.; John Wiley & Sons, Inc., 2003. (b) Matthews, V.; Kubler, D. J. Org. Chem. 1960, 25, 266–268.

(11) (a) Lyttle, D. A.; Weisblat, D. I. J. Am. Chem. Soc. 1947, 69, 2118–2119. (b) Snyder, B. H. R.; Katz, L. J. Am. Chem. Soc. 1947, 69, 3140–3142. (c) Genet, J. P.; Ferround, D. Tetrahedron Lett. 1984, 25, 3579–3582. (d) Fiandanese, V.; Naso, F.; Scilimati, A. Tetrahedron Lett. 1984, 25, 1187–1190. (e) Metz, A. E.; Berritt, S.; Dreher, S. D.;

Kozlowski, M. C. Org. Lett. 2012, 14, 760-763.
(12) (a) Kornblum, N.; Chalmers, M. E.; Daniels, R. J. Am. Chem. Soc. 1955, 77, 6654-6655. (b) Kornblum, N.; Blackwood, R. K.; Powers, J. W. J. Am. Chem. Soc. 1957, 79, 2507-2509.

(13) Kornblum, N.; Weaver, W. M. J. Am. Chem. Soc. 1958, 80, 4333-4337.

(14) Simchen, G. Liebigs Ann. Chem. 1979, 1979, 628-638.

(15) Carmeli, M.; Rozen, S. J. Org. Chem. 2006, 71, 4585-4589.

(16) Sifniades, S. J. Org. Chem. 1975, 40, 3562-3566.

(17) Kislyi, V. P.; Laikhter, A. L.; Ugrak, B. I.; Semenov, V. V. Russ. Chem. Bull. 1994, 43, 70-74.

(18) Laikhter, A. L.; Kislyi, V. P.; Semenov, V. V. Mendeleev Commun. 1993, 3, 20–21.

(19) Nakaike, Y.; Taba, N.; Itoh, S.; Tobe, Y.; Nishiwaki, N.; Ariga, M. Bull. Chem. Soc. Jpn. **200**7, 80, 2413–2417.

(20) Hessel, V.; Kralisch, D.; Kockmann, N.; Noël, T.; Wang, Q. ChemSusChem 2013, 6, 746-789.

(21) Kulkarni, A. A. Beilstein J. Org. Chem. 2014, 10, 405-424.

(22) (a) Brocklehurst, C. E.; Lehmann, H.; La Vecchia, L. Org. Process Res. Dev. 2011, 15, 1447–1453. (b) Yu, Z.; Zhou, P.; Liu, J.;

Wang, W.; Yu, C.; Su, W. Org. Process Res. Dev. 2016, 20, 199–203.
(23) See Supporting Information for details. The product yield does

not depend on the duration of the stirring with desiccant, even after 3 h stirring of the reaction mixture with magnesium sulfate the yield of the reaction did not change.

(24) Yield determined by 1H NMR using mesytline as internal standard due to the volatility of the product.

(25) See Experimental Section for details

(26) Simmons, T.; Kreuz, K. J. Org. Chem. 1968, 33, 836-837.

(27) See Experimental Section for details

(28) Pinnick, H. W. In Organic Reactions; John Wiley & Sons, Inc., 2004.

(29) Gottlieb, H. E.; Kotlyar, V.; Nudelman, A. J. Org. Chem. 1997, 62, 7512-7515.

(30) Zen, S.; Kaji, E. Org. Synth. 1977, 57, 60.

(31) Niyazymbetov, M. E.; Evans, D. H. J. Org. Chem. **1993**, 58, 779–783.

(32) Kim, S.-U.; Liu, Y.; Nash, K. M.; Zweier, J. L.; Rockenbauer, A.; Villamena, F. A. J. Am. Chem. Soc. **2010**, 132, 17157–17173.

(33) Feuer, H.; Monter, R. P. J. Org. Chem. 1969, 34, 991–995.

(34) (a) Martynov, I. V.; Zavel'skii, V. O.; Kovalenko, S. V.; Yurtanov, A. I. *Izv. Akad. Nauk SSSR, Ser. khim.* **1982**, 1029–1033.

(b) Martynov, I. V.; Zavel'skii, V. O.; Kovalenko, S. V.; Fetisov, V. I.;

Yurtanov, A. I. Dokl. Akad. Nauk SSSR 1983, 152-156.

(35) Sin, I.; Kang, C. S.; Bandara, N.; Sun, X.; Zhong, Y.; Rogers, B. E.; Chong, H.-S. *Bioorg. Med. Chem.* **2014**, *22*, 2553–2562.

(36) Takeuchi, Y.; Itoh, N.; Koizumi, T.; Yamagami, C.; Takeuchi, Y. *Magn. Reson. Chem.* **1992**, 30, 58–64.

(37) Paine, J. B.; Brough, J. R.; Buller, K. K.; Erikson, E. E.; Dolphin, D. J. Org. Chem. **1987**, *52*, 3993–3997.

(38) Sedelmeier, J.; Ley, S. V.; Baxendale, I. R.; Baumann, M. Org. Lett. 2010, 12, 3618-3621.

(39) Greene, B.; Lewis, K. Aust. J. Chem. 1968, 21, 1845-1852.

(40) Schröder, K.; Join, B.; Amali, A. J.; Junge, K.; Ribas, X.; Costas, M.; Beller, M. Angew. Chem., Int. Ed. 2011, 50, 1425–1429.